

Advanced preclinical *in vitro* and *in vivo* characterization of a novel, non-hydroxamate-based LpxC inhibitor for the intravenous and oral treatment of multidrug-resistant Enterobacterales

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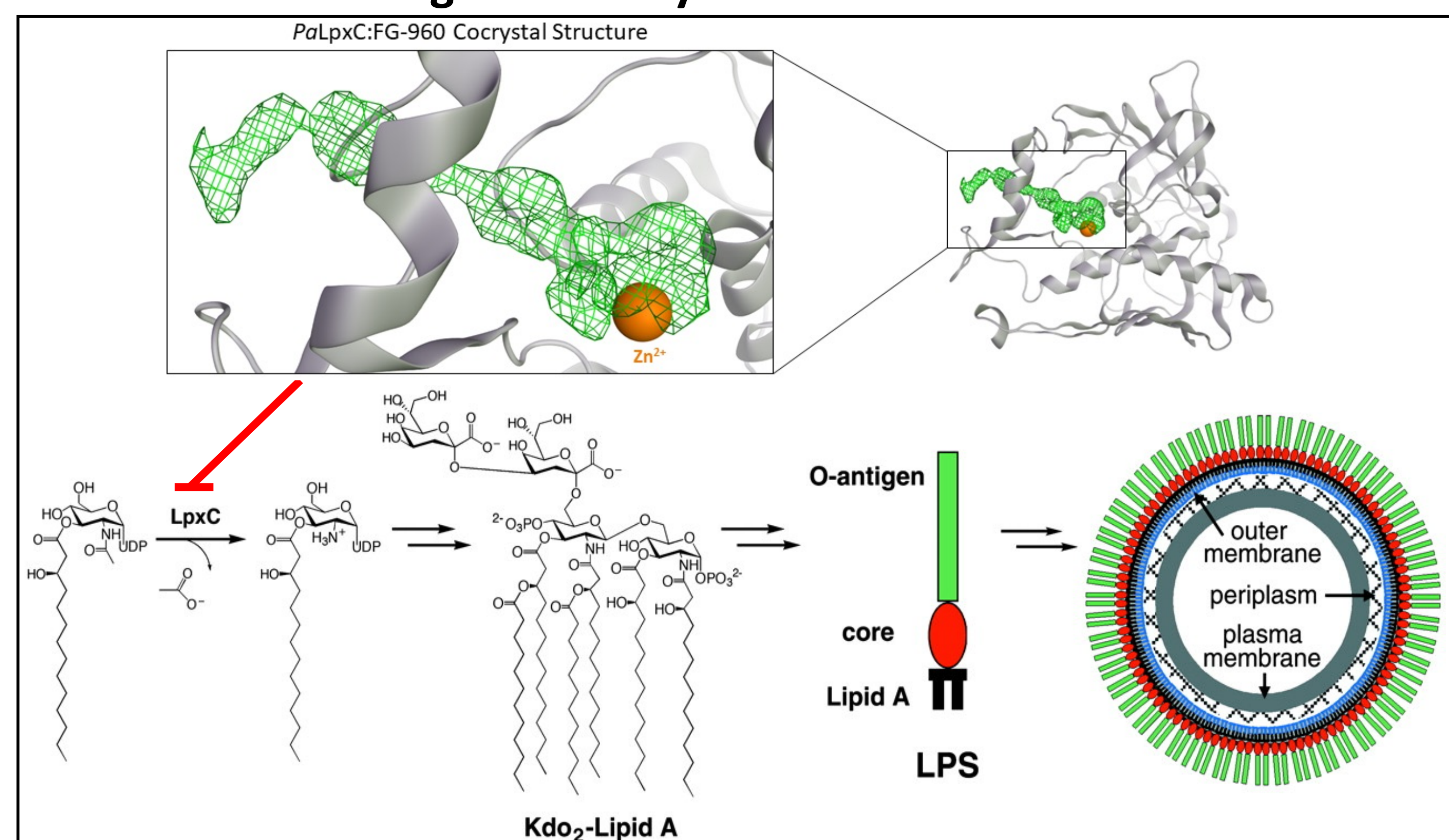
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Abstract

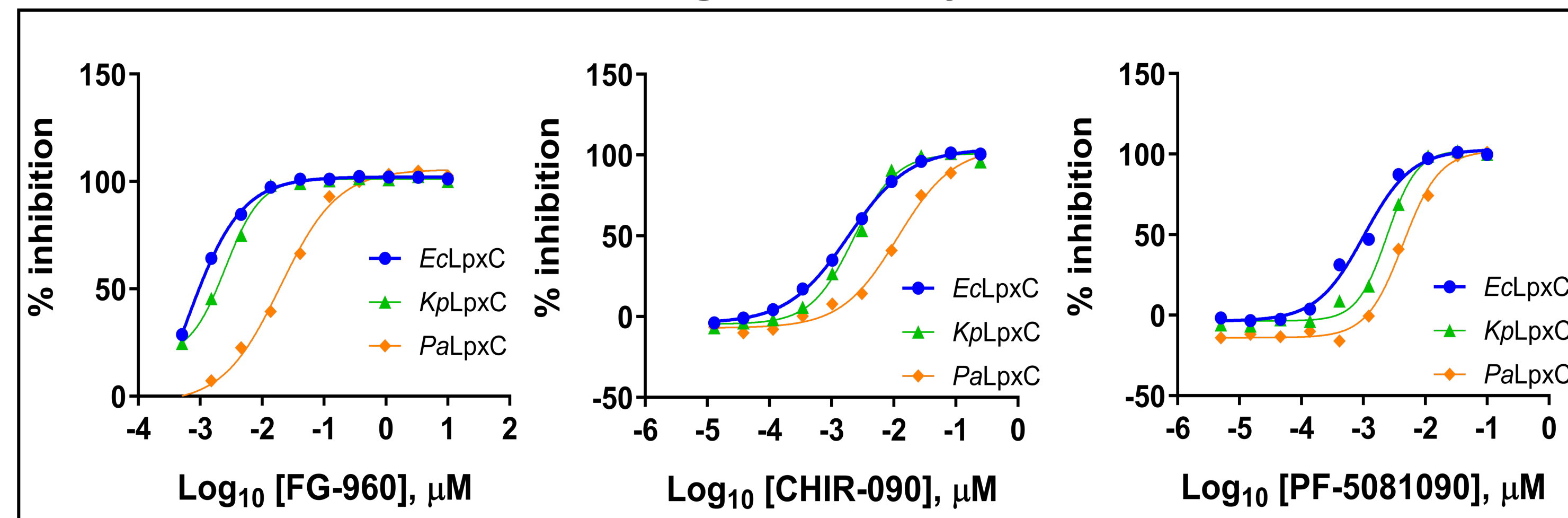
LpxC is an essential metalloenzyme that Gram-negative bacteria require for outer membrane biosynthesis. Previous antibacterial efforts have focused on hydroxamate-based small molecules to engage LpxC's catalytic zinc. Unfortunately, these previous attempts have been hampered by safety liabilities. Leveraging its metalloenzyme inhibitor platform, Blacksmith has identified FG-960, a non-hydroxamate-based LpxC inhibitor with strong *in vitro* and *in vivo* profiles against MDR Enterobacterales, and is currently being advanced as a treatment for urinary tract infections (UTIs).

FG-960 is highly-potent against the LpxC enzyme (*E. coli* IC₅₀ = 1 nM), and has strong whole cell activity against panels of contemporary MDR and non-MDR Enterobacterales isolates (MIC₉₀ = 4 µg/ml; n = 973 strains). Static time kill studies demonstrate FG-960's rapid bactericidal activity, with regrowth suppression evident at concentrations ≥2X MIC. When evaluated in standard frequency of resistance assays, FG-960 shows spontaneous resistance emergence at frequencies consistent with historical LpxC inhibitors (1x10⁻⁸-1x10⁻¹⁰) at 4X MIC against multiple MDR and non-MDR strains. Pharmacokinetic studies have demonstrated dose-proportional exposures in the plasma from preclinical species, with considerable accumulation of intact FG-960 in the urinary tract. This higher localized exposure likely contributes to FG-960's strong *in vivo* efficacy profile, with >1-log kill achieved using either intravenous or oral administration at dose levels of 10-50 mg/kg/day in a mouse UTI model. Importantly, and in multiple preclinical models, FG-960 does not exhibit cardiovascular toxicity observed with previous hydroxamate-based LpxC inhibitors. In summary, Blacksmith is advancing FG-960 in ongoing IND-enabling studies for the treatment of UTIs caused by MDR Gram-negative bacteria.

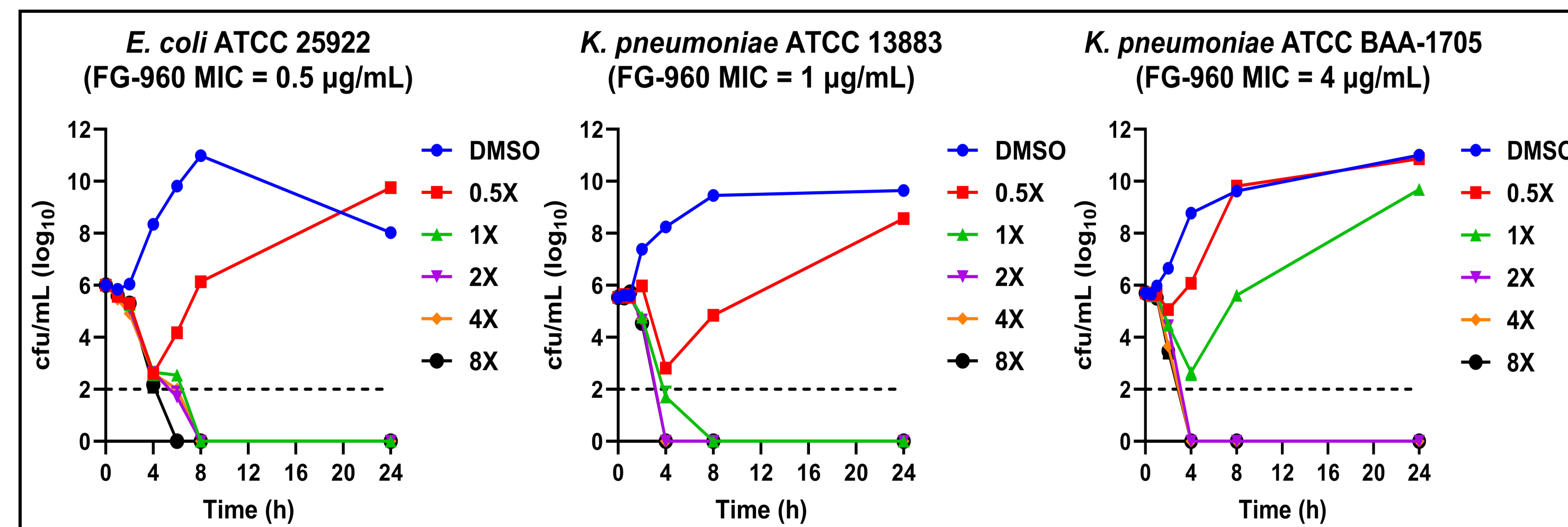
Target Pathway and FG-960 MOA



Target Potency



In vitro Kill Kinetics



In vitro static time-kill results with FG-960 evaluated against three different Enterobacterales target strains. Compound was added to exponentially-growing cultures of each indicated isolate at the fold-MIC concentrations specified. Recoverable bioburdens were quantified via viable plate counting at 0, 1, 2, 4, 6, 8, and 24 hours post-compound addition.

In vitro Activity Spectrum (MIC₉₀)

Antimicrobial agent	<i>E. coli</i> (n=108)		<i>K. pneumoniae</i> (n=107)		<i>K. oxytoca</i> (n=56)		<i>K. aerogenes</i> (n=52)		<i>P. mirabilis</i> (n=54)	
	MIC ₅₀	MIC ₉₀	MIC ₅₀	MIC ₉₀	MIC ₅₀	MIC ₉₀	MIC ₅₀	MIC ₉₀	MIC ₅₀	MIC ₉₀
FG-960	0.5	1	0.5	4	1	2	0.5	2	1	4
Ceftazidime-avibactam	0.06	0.25	0.12	0.25	0.12	0.25	0.12	0.25	≤0.03	0.06
Ceftriaxone	0.06	>8	0.06	>8	0.06	>8	0.12	>8	≤0.004	0.008
Ciprofloxacin	≤0.06	64	≤0.06	64	≤0.06	0.25	≤0.06	0.12	≤0.06	4
Meropenem	0.015	0.03	0.03	0.06	0.03	0.03	0.03	0.06	0.12	0.12
Amoxicillin-clavulanate	8	16	2	16	2	16	32	>32	1	8
Nitrofurantoin	16	32	64	>64	32	64	64	>64	>64	>64
Piperacillin-tazobactam	2	16	4	32	2	128	4	32	0.25	1
Trimethoprim-sulfamethoxazole	≤0.12	>16	≤0.12	>16	≤0.12	4	≤0.12	0.25	≤0.12	>16

Antimicrobial agent	<i>C. freundii</i> (n=52)		<i>C. koseri</i> (n=54)		<i>E. cloacae</i> (n=52)		<i>M. morgani</i> (n=50)		<i>S. marcescens</i> (n=51)	
	MIC ₅₀	MIC ₉₀	MIC ₅₀	MIC ₉₀	MIC ₅₀	MIC ₉₀	MIC ₅₀	MIC ₉₀	MIC ₅₀	MIC ₉₀
FG-960	1	4	0.5	1	1	2	2	4	0.5	1
Ceftazidime-avibactam	0.12	0.5	0.06	0.12	0.25	1	0.06	0.25	0.25	0.5
Ceftriaxone	0.25	>8	0.06	0.12	0.5	>8	0.06	4	0.5	>8
Ciprofloxacin	≤0.06	4	≤0.06	≤0.06	≤0.06	0.5	≤0.06	2	≤0.06	2
Meropenem	0.03	0.06	0.015	0.03	0.03	1	0.06	0.12	0.06	0.12
Amoxicillin-clavulanate	32	>32	2	4	>32	>32	>32	>32	>32	>32
Nitrofurantoin	32	32	64	>64	64	>64	64	>64	>64	>64
Piperacillin-tazobactam	4	128	2	4	4	>128	0.25	128	2	32
Trimethoprim-sulfamethoxazole	≤0.12	>16	≤0.12	≤0.12	≤0.12	>16	≤0.12	>16	0.5	4

MIC profiling of FG-960 against contemporary and geographically-distinct clinical isolates of Enterobacterales pathogens commonly found to cause UTIs.

Frequency and Mechanism of FG-960 Resistance

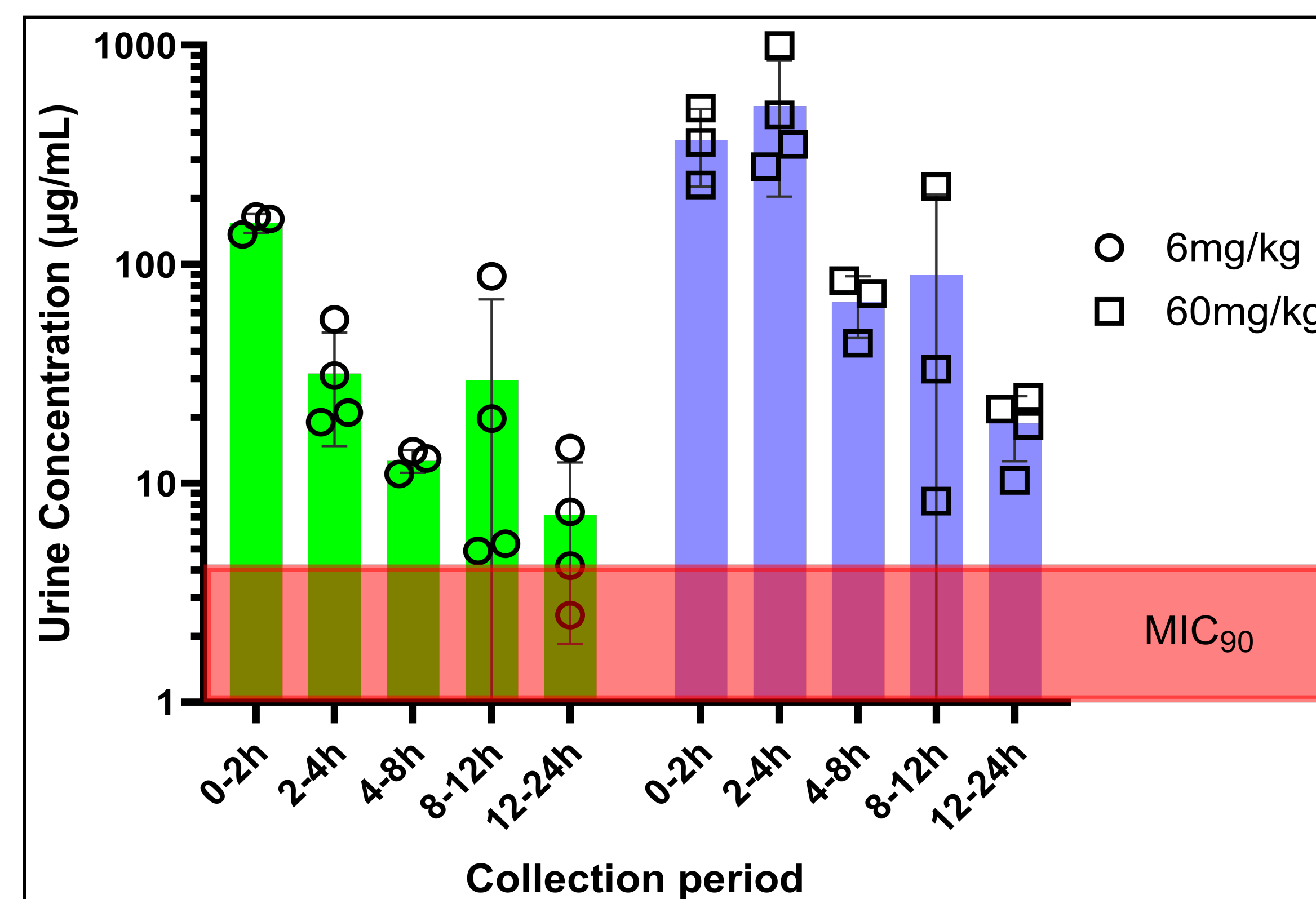
Compound	Fold MIC	<i>E. coli</i> ATCC 25922		<i>K. pneumoniae</i> ATCC 13883	
		FoR ^a	Fold-Change (vs WT)	FoR ^a	Fold-Change (vs WT)
FG-960	4X	1.83x10 ⁻¹⁰	NA	1.84x10 ⁻⁸	NA
	8X	<1.83x10 ⁻¹⁰	NA	1.35x10 ⁻⁹	NA
	16X	<1.83x10 ⁻¹⁰	NA	<2.70x10 ⁻¹⁰	NA
ACHN-975	4X	9.14x10 ⁻¹⁰	NA	2.05x10 ⁻⁸	NA
	8X	<1.83x10 ⁻¹⁰	NA	2.70x10 ⁻⁹	NA
	16X	<1.83x10 ⁻¹⁰	NA	<2.70x10 ⁻¹⁰	NA
PF-5081090	4X	1.33x10 ⁻⁸	NA	Not Tested	NA
	8X	1.33x10 ⁻⁸	NA	Not Tested	NA
	16X	3.66x10 ⁻¹⁰	NA	Not Tested	NA
Ciprofloxacin	4X	1.10x10 ⁻⁹	NA	Not Tested	NA
	8X	1.10x10 ⁻⁹	NA	Not Tested	NA
	16X	<1.83x10 ⁻¹⁰	NA	Not Tested	NA

Strain	Sequencing Results		MIC (µg/mL)								
	LpxC	FabZ	FG-960	Fold-Change (vs WT)	CIP ^a	Fold-Change (vs WT)	TOB ^b	Fold-Change (vs WT)	MPM ^c	Fold-Change (vs WT)	
<i>E. coli</i> 25922	WT	WT	0.25	NA	0.016	NA	0.5	NA	0.016	NA	
<i>E. coli</i> ATCC 25922 clones from FG-960 FoR ^a plates	#1	WT	A71S	2	8	0.008	2	0.5	1	0.016	1
	#2	WT	A71V	2	8	0.008	2	0.25	0.5	0.008	0.5
	#3	WT	K102N	4	16	0.008	2	0.5	1	0.016	1
	#4	WT	R100S	8	32	0.008	2	0.125	0.25	≤0.008	≤0.5
	#5	WT	C139Y	2	8	0.008	2	0.5	1	0.016	1
	#6	WT	P22L	2	8	0.008	2	0.25	0.5	≤0.008	≤0.5
	#7	WT	G129E	2	8	0.008	2	0.125	0.25	≤0.008	≤0.5
	#8	WT	A71S	4	16	0.016	1	0.5	1	0.016	1

TOP: Spontaneous frequency of resistance (FoR) results for FG-960, two comparator LpxC inhibitors, and ciprofloxacin evaluated against *E. coli* ATCC 25922 and *K. pneumoniae* ATCC 13883.

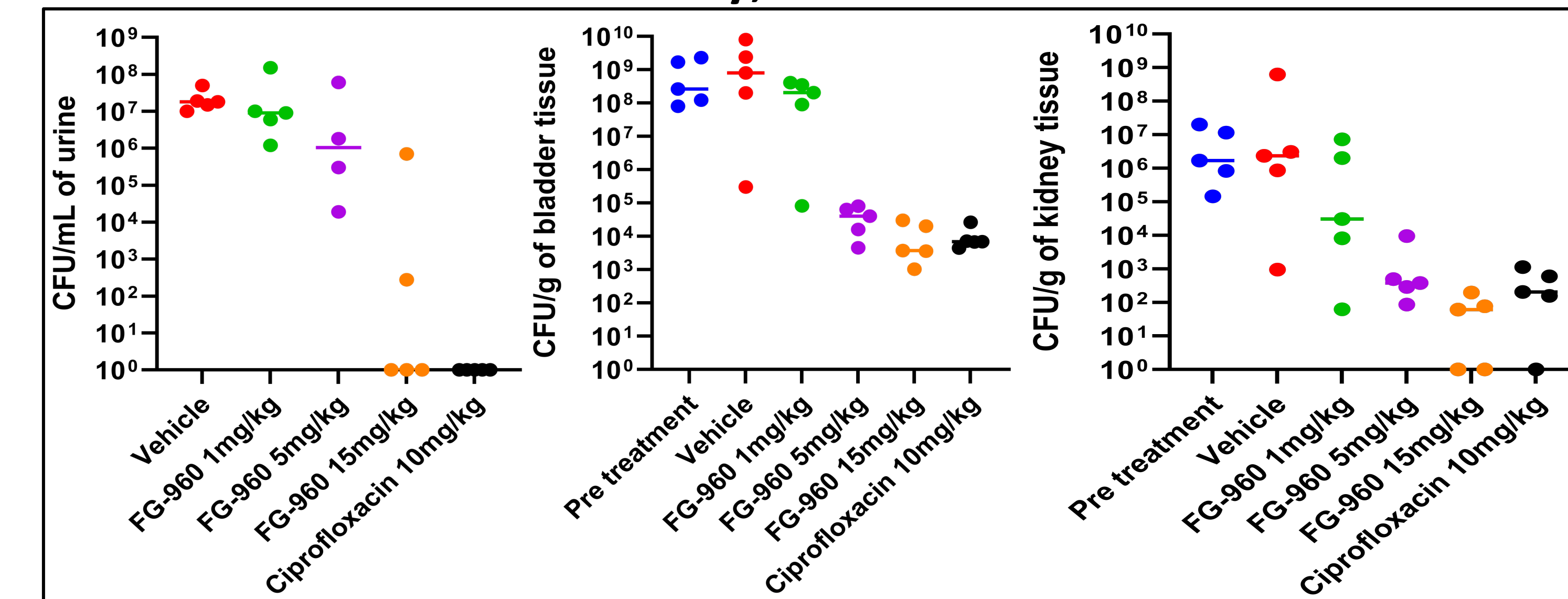
BOTTOM: Sequencing and antibiogram phenotyping of select *E. coli* ATCC 25922 spontaneous-resistant mutants recovered after FG-960 challenge in FoR studies.

In vivo Pharmacokinetics



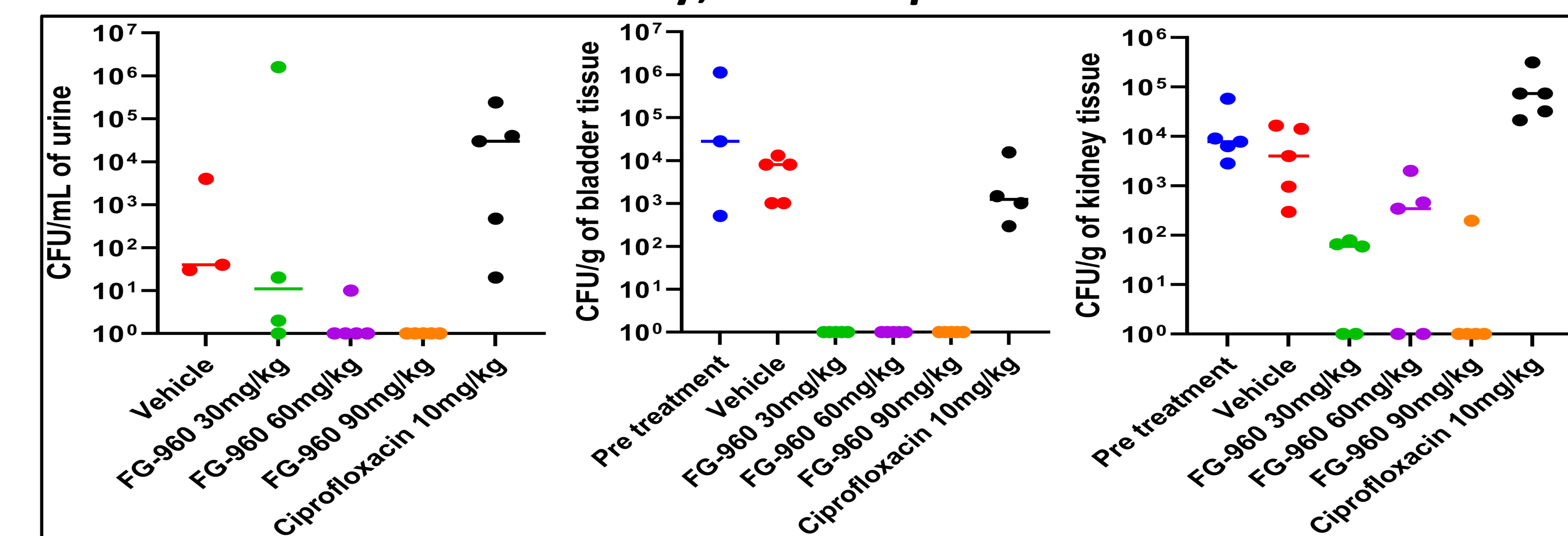
Kinetic recovery of intact FG-960 from Sprague Dawley rat urine (n=4) after IV administration of 6 mg/kg (green bars, circles) and 60 mg/kg (blue bars, squares) doses. Urine was recovered via metabolic cage collection. For reference, the anticipated MIC₉₀ for all target uropathogens is shaded in red.

IV-efficacy; non-MDR *E. coli*



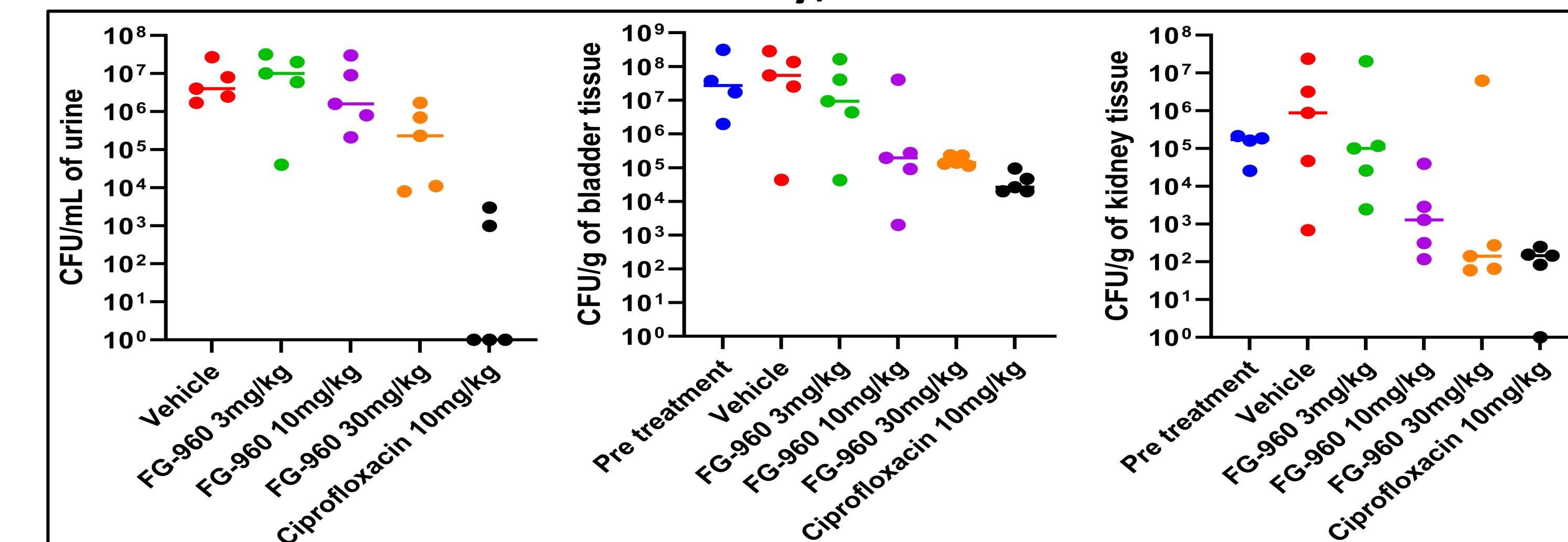
E. coli UTI89 (FG-960 MIC = 0.5 µg/mL) was introduced via transurethral administration 24 hours prior to initiation of therapy. FG-960 was administered BID for 72 hours prior to bioburden recoveries from the urine (left), bladder (middle), or kidney (right). Ciprofloxacin was included as a comparator.

IV-efficacy; MDR *K. pneumoniae*



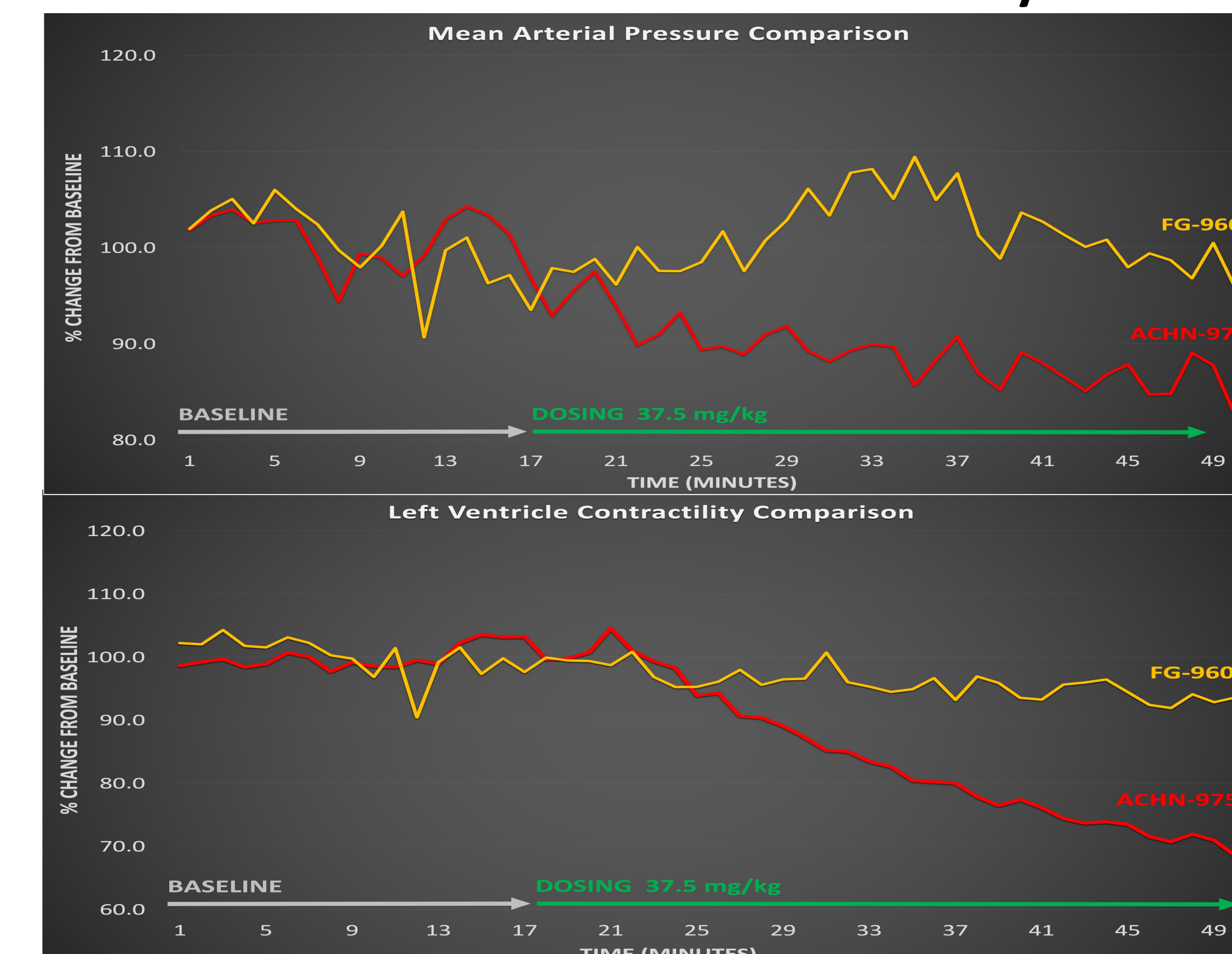
K. pneumoniae BAA-1705 (FG-960 MIC = 4 µg/mL), a blaKPC-2-producing and fluoroquinolone-resistant clinical isolate, was introduced via transurethral administration 24 hours prior to initiation of therapy. FG-960 was administered QID for 72 hours prior to bioburden recoveries from the urine (left), bladder (middle), or kidney (right). Ciprofloxacin was included as a comparator.

PO-efficacy; MDR *E. coli*



E. coli NCTC 13462 (FG-960 MIC = 0.25 µg/mL), a blaCTX-M-2-producing clinical isolate, was introduced via transurethral administration 24 hours prior to initiation of therapy. FG-960 was administered BID for 72 hours prior to bioburden recoveries from the urine (left), bladder (middle), or kidney (right). Ciprofloxacin was included as a comparator.

Preclinical Cardiovascular Safety



Anesthetized rat model data demonstrating differentiation of FG-960 (yellow) from ACHN-975 (red). Both mean arterial pressure (top panel) and left ventricle contractility (bottom panel) were reduced both preclinically and in a Phase 1 clinical trial by ACHN-975, but no significant changes in either parameter were observed when FG-960 was administered at 37.5 mg/kg.

Acknowledgements

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