

# Small molecule inhibitor of FEN1 nuclease utilizing a novel metal-binding pharmacophore synergizes with inhibitors of USP1, PARG, PARG and ATR

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## Abstract

Flap endonuclease 1 (FEN1) is a structure-specific metallo nuclease that cleaves 5' DNA flaps during replication and repair. FEN1 is an attractive target for development of anticancer therapeutics because it is overexpressed in many tumor types and has a large number of synthetic lethality partners including genes in Homologous Recombination (HR) pathway (Mengwasser et al., 2019; Guo et al., 2020).

Utilizing fragment-based drug discovery (FBDD) approach we identified a novel metal-binding pharmacophore that binds to the two magnesium ions in the FEN1 active site. Further elaboration using fragment growth strategies resulted in highly potent and selective inhibitors. The current lead (BSM-1516) is ~65-fold more potent against FEN1 than its related enzyme Exonuclease 1 (Exo1) in biochemical assays (IC50 of 7 nM and 460 nM, respectively), an improvement of more than an order of magnitude in selectivity compared to earlier efforts. FEN1 target engagement in live cells was validated by cellular thermal shift assay (CETSA™ EC50 of 24 nM). Inhibition of FEN1 led to its increased association with chromatin in S-phase cells and recruitment of PARP1 enzyme.

In clonogenic assay, BRCA2-deficient DLD1 cells were ~15-fold more sensitive to FEN1 inhibition than their isogenic BRCA2-wild-type counterparts (EC50 of 350 nM and 5 μM, respectively), confirming the increased susceptibility of HR deficient cancer cells to FEN1 inhibition. Treatment of BRCA2-deficient but not wild-type DLD1 cells with BSM-1516 resulted in cell cycle arrest and dose-dependent decrease of S-phase BrdU incorporation into DNA. Cell cycle arrest of BRCA2-deficient DLD1 cells was accompanied by DNA damage signaling and by accumulation of chromatin-bound RPA32, a marker of ssDNA.

To explore synthetic lethal interactions of FEN1 with other DNA repair genes we performed FEN1-inhibitor-anchored CRISPR screen. This analysis revealed that in addition to HR pathway inactivation, genomic perturbations in EXO1, USP1 and PARP1 genes sensitized cells to FEN1 inhibition. Synergistic relationships of BSM-1516 and its combination potential were further explored in viability studies with a panel of DDR inhibitors (n=25) in BRCA2-proficient and deficient cell lines. Strong synergy was identified with multiple drug classes that included inhibitors of USP1 (KSQ-4279), PARP (Olaparib, Niraparib, Talazoparib, AZD5305), PARG (PDD 00017273) and ATR (AZD6738, VE-822, Elimusertib).

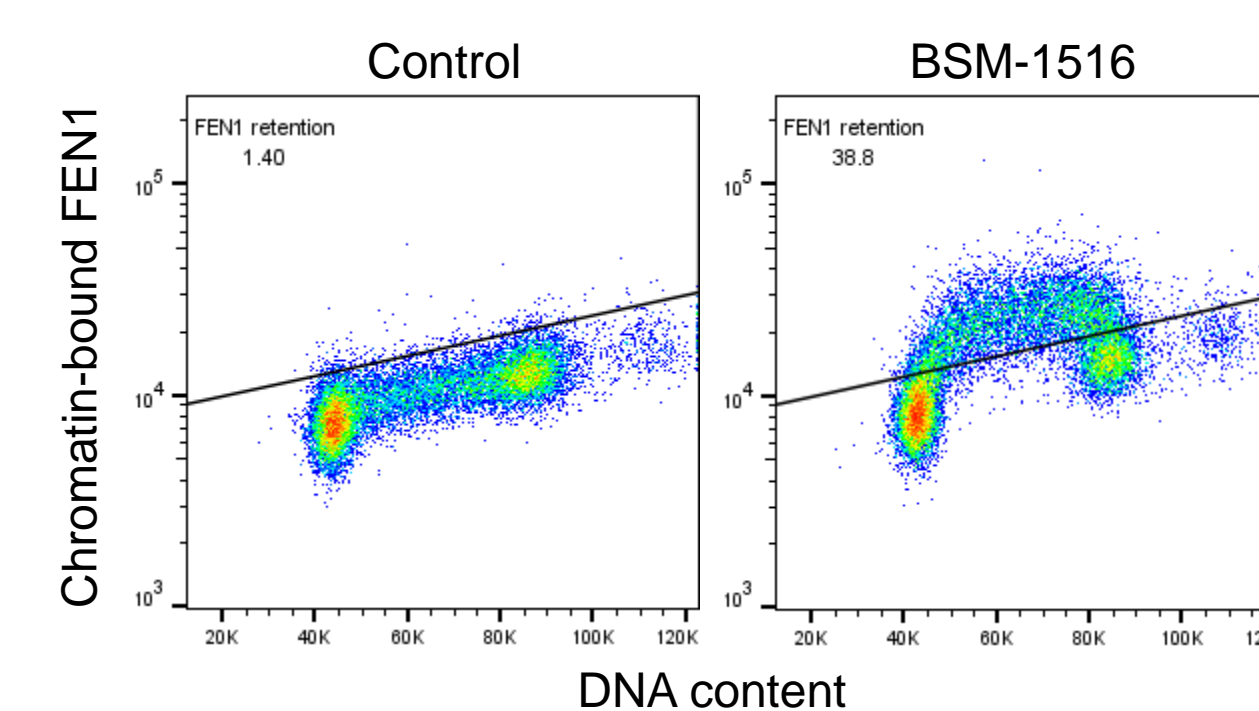
In vitro ADME assays and in vivo PK studies showed that BSM-1516 has properties suitable for in vivo testing, either as a single agent or in combination with synergistic DDR inhibitors, an investigation that is currently underway.

## Cellular Target Engagement and Mechanism of Action

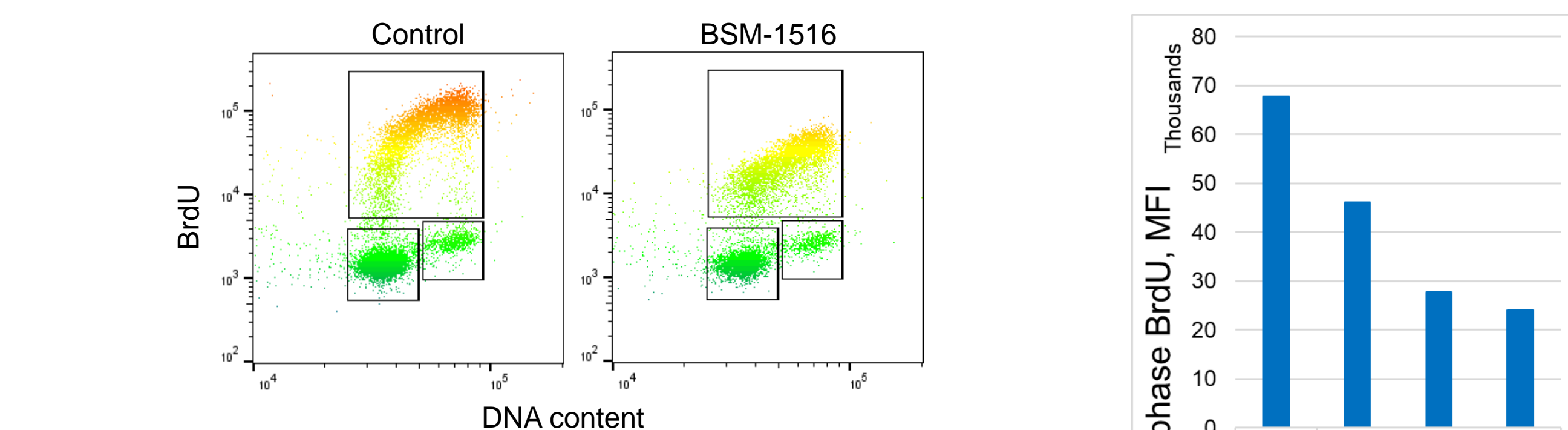
BSM-1516 stabilizes FEN1 and increases its chromatin retention in S phase that is accompanied by slowdown of DNA replication and Talazoparib-mediated PARP1 trapping.

Company	ID	CETSA EC50 (μM)
Astra Zeneca	Cmpd 4	1.2
Athersys	Cmpd 8	0.21
Ideaya	Cmpd 12	>100
Blacksmith Medicines	BSM-1516	0.024

Cellular thermal shift assay (CETSA™) was performed using InCELL Pulse™ target engagement assay (DiscoverX) in stably transfected HEK293 cells expressing FEN1 catalytic domain with C-terminal ePL tag.

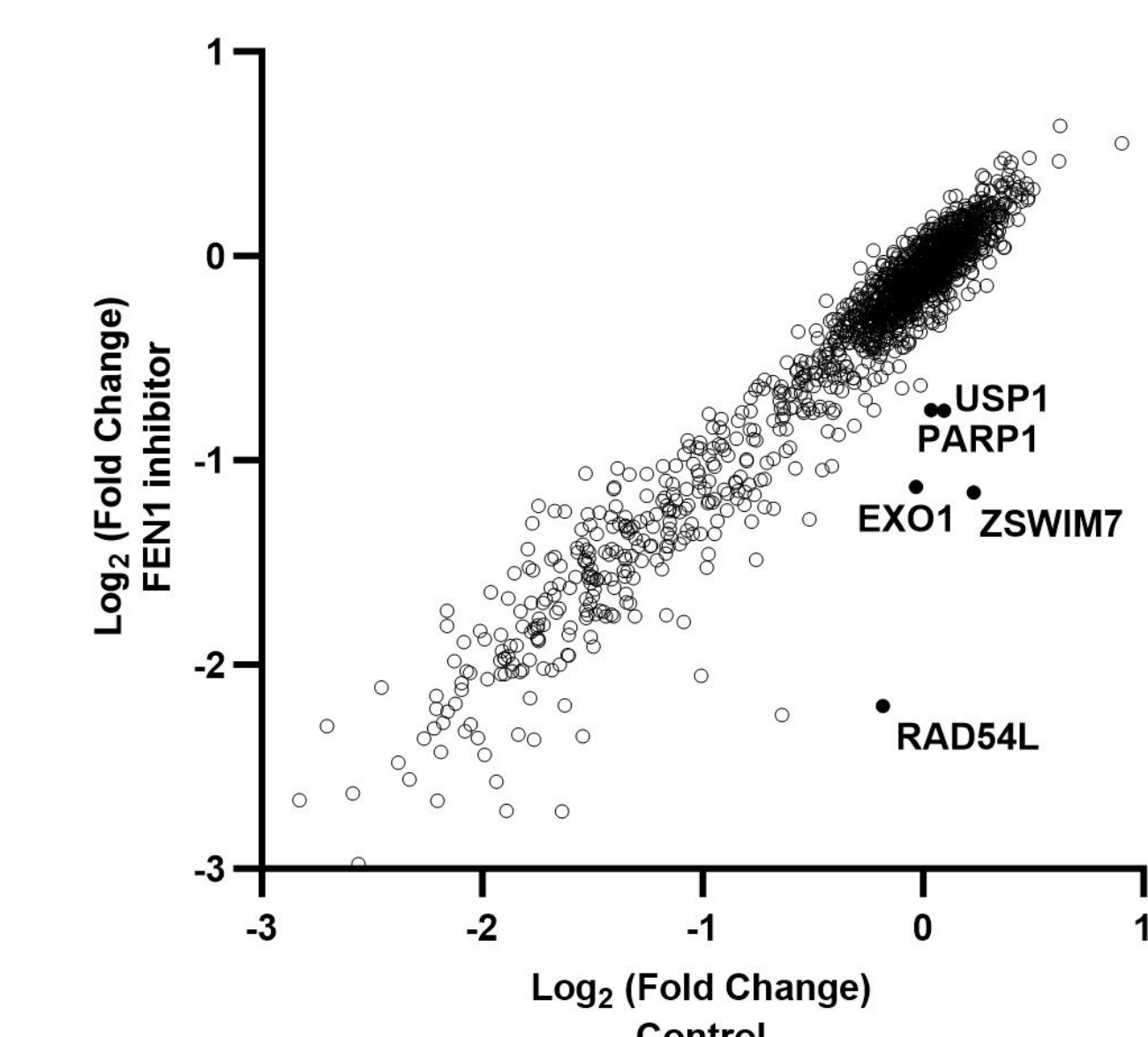


Intracellular staining of endogenous chromatin-bound FEN1 in DLD1 cells treated with 3μM BSM-1516 for 16h. Soluble FEN1 was extracted with detergent prior to fixation and subsequent analysis by flow cytometry.



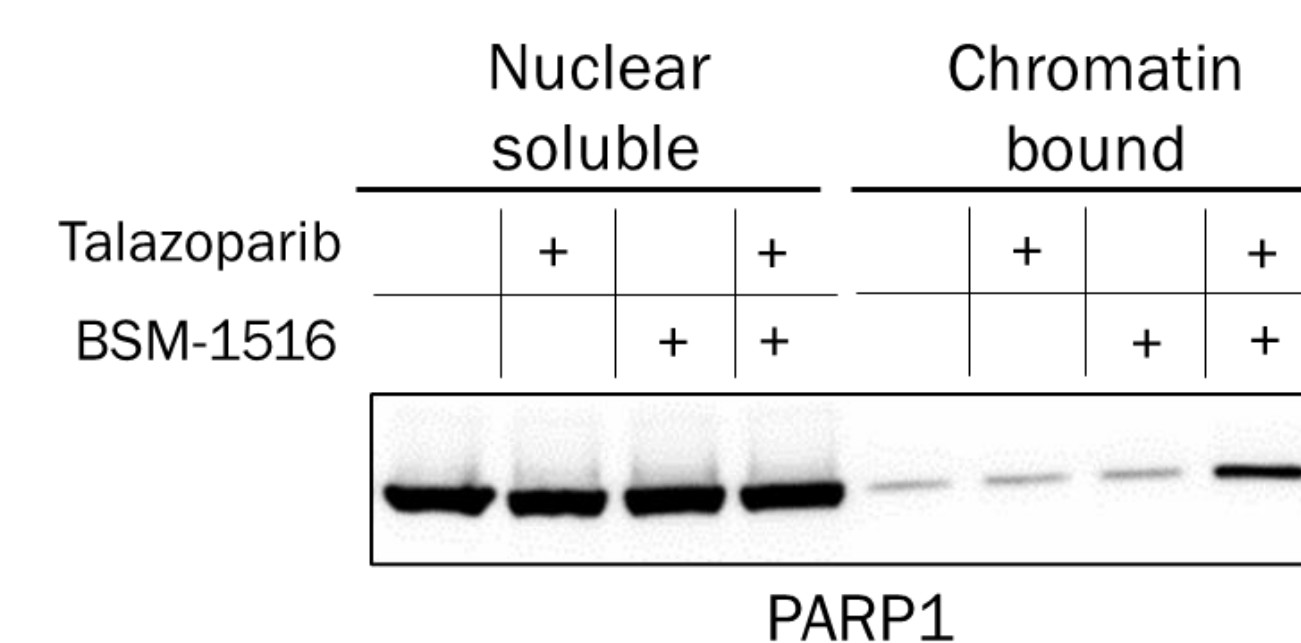
BSM-1516 causes replication suppression and impaired S phase progression. Analysis of BrdU incorporation and total DNA content monitored by flow-cytometry after 16h of BSM-1516 treatment.

## CRISPR screen identifies genes that are synthetic lethal with FEN1 inhibition

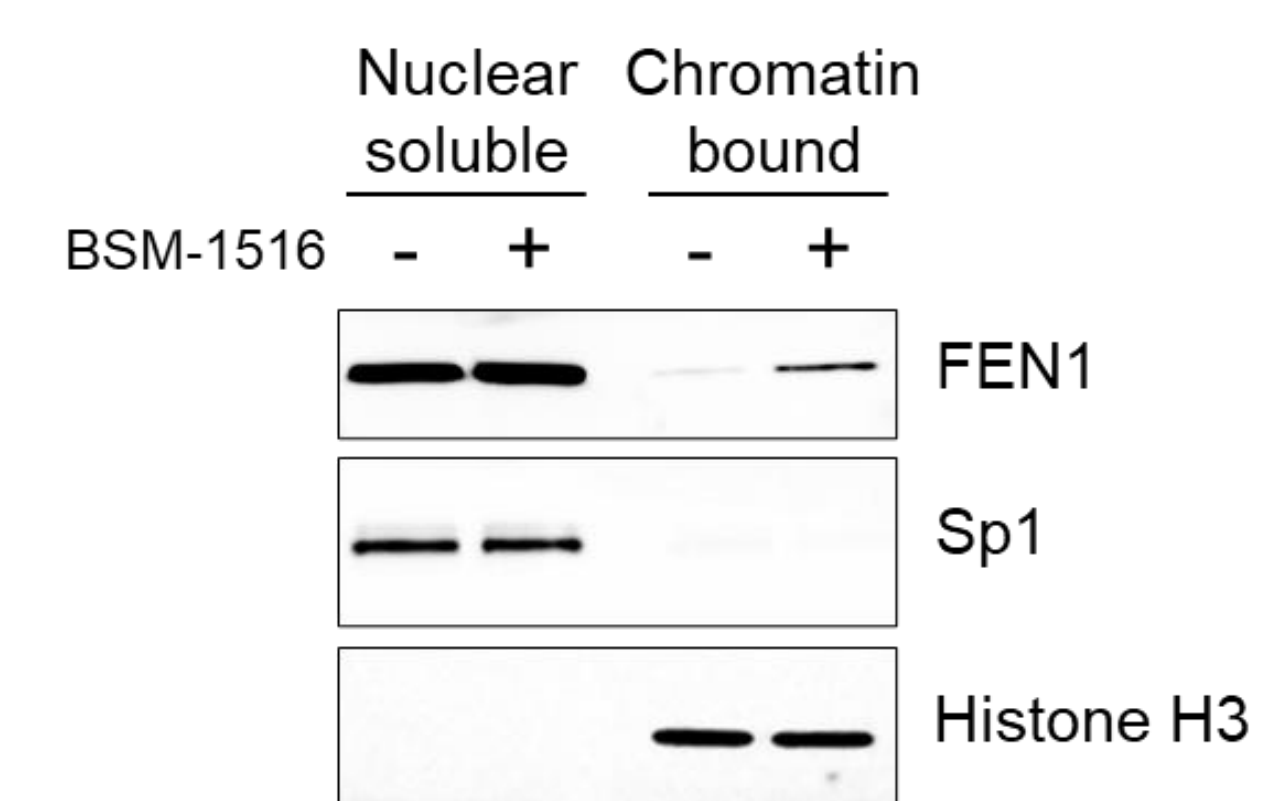


To identify genetic determinants driving FEN1 inhibitor sensitivity we performed CRISPR-Cas9 loss-of-function screen using lentiviral sgRNA sublibrary that targets DNA damage response genes and metalloenzymes. Data on the graph were plotted as ratio of sgRNA abundance on day 14 versus day 0 for control and for FEN1-inhibitor (2μM) treated DLD1 cells.

Our screen confirmed several previously published FEN1 gene interactors including USP1 (Fielden et al., 2023 bioRxiv), PARP1 (Hanzlikova et al., 2018), EXO1 (Tishkoff et al., 1997) and homologous recombination repair (HRR) pathway defects (Mengwasser et al., 2019; Guo et al., 2020). PARG and WDR48 genes were not targeted by the library.



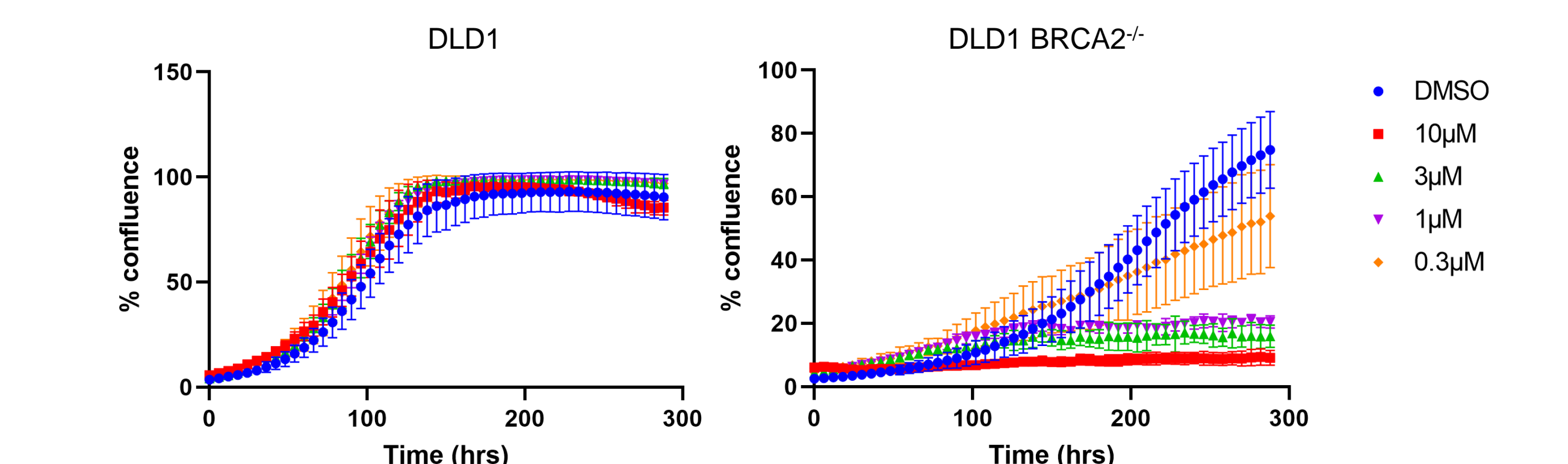
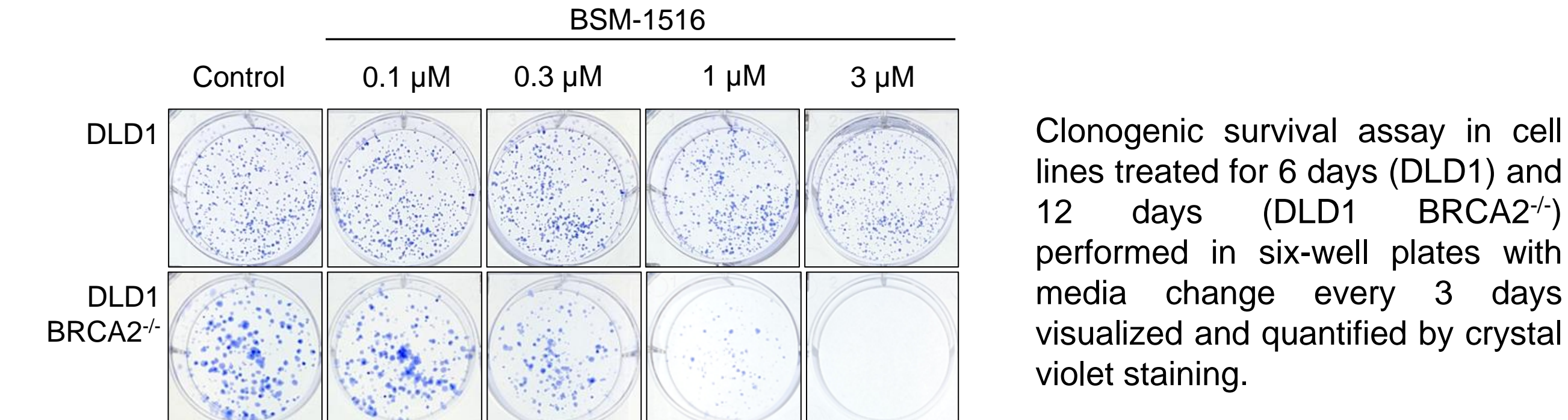
Western blot analysis of endogenous chromatin-bound PARP1 in DLD1 cells treated with indicated inhibitors for 3h and then fractionated.



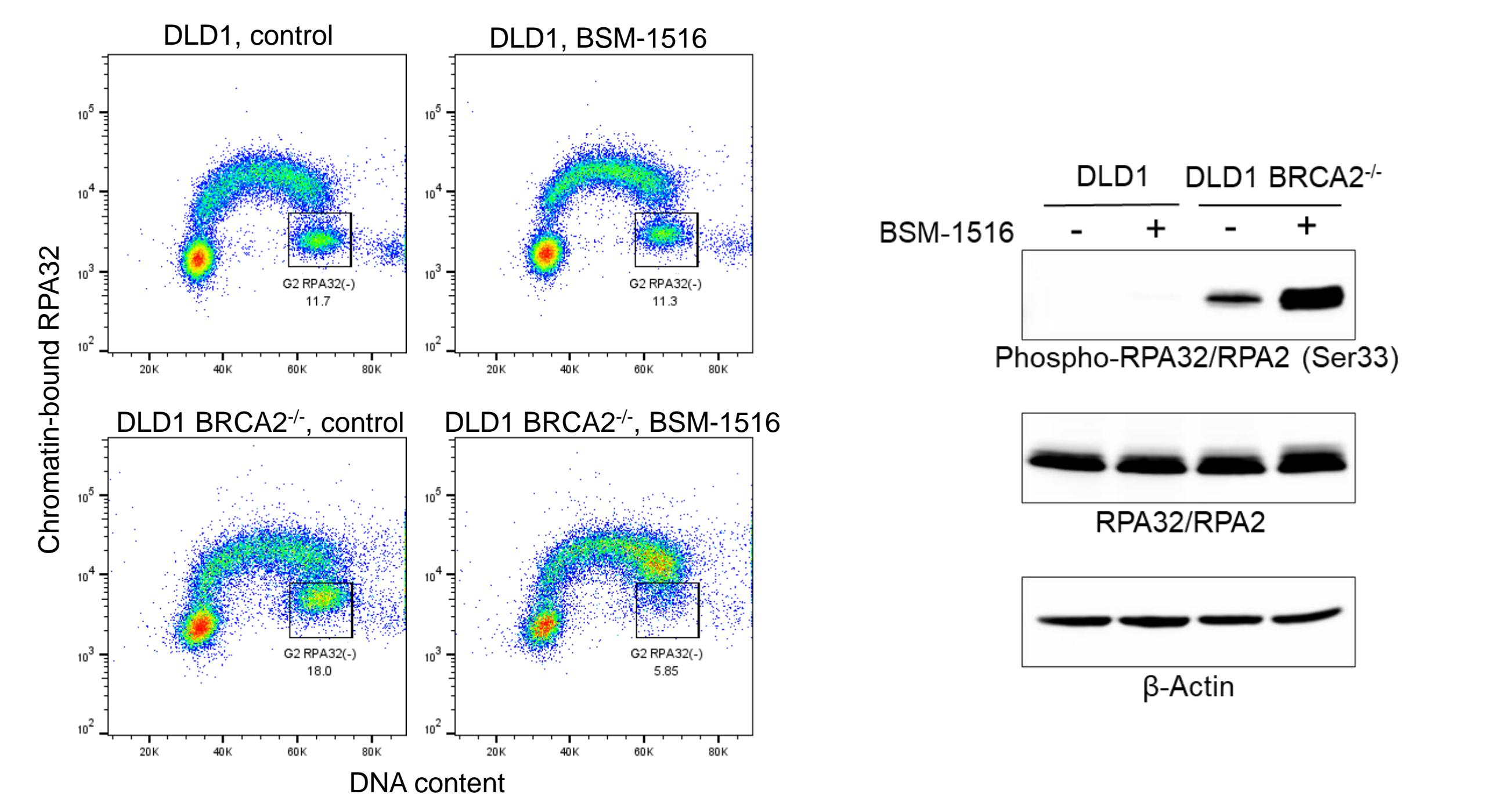
Western blot analysis of endogenous chromatin-bound FEN1 in S phase. DLD1 cells were arrested by double thymidine block, released for 3h into fresh media with or without 3μM BSM-1516 and then fractionated.

## Viability assays: BRCA2-deficient cells are >10 fold more sensitive to FEN1 inhibitor than isogenic wild-type control

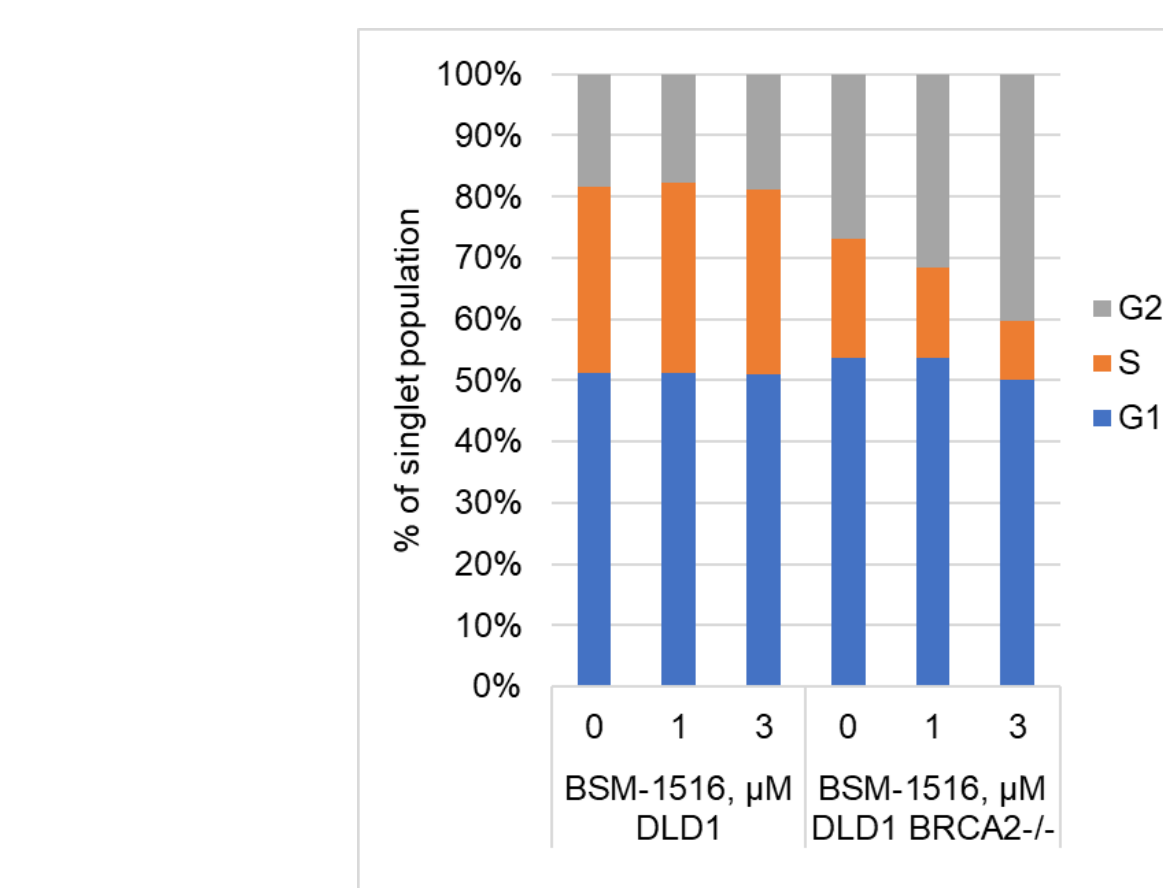
Consistent with the previous reports (Mengwasser et al., 2019; Guo et al., 2020) BRCA2 gene deficiency causes sensitivity to pharmacologic inhibition of FEN1. In cell survival and culture confluency assays DLD1 cells carrying a truncating mutation of BRCA2 gene were more than 10-fold more sensitive to FEN1 inhibition than BRCA2-proficient isogenic control cells. Additionally, BRCA2-deficient cells show higher levels of RPA-coated ssDNA in G2, RPA S33 phosphorylation and G2/M arrest in response to BSM-1516 treatment.



Real-time cell proliferation growth curves were generated by confluence imaging every 6 hours with triplicate replicates on Incucyte live-cell analysis system.



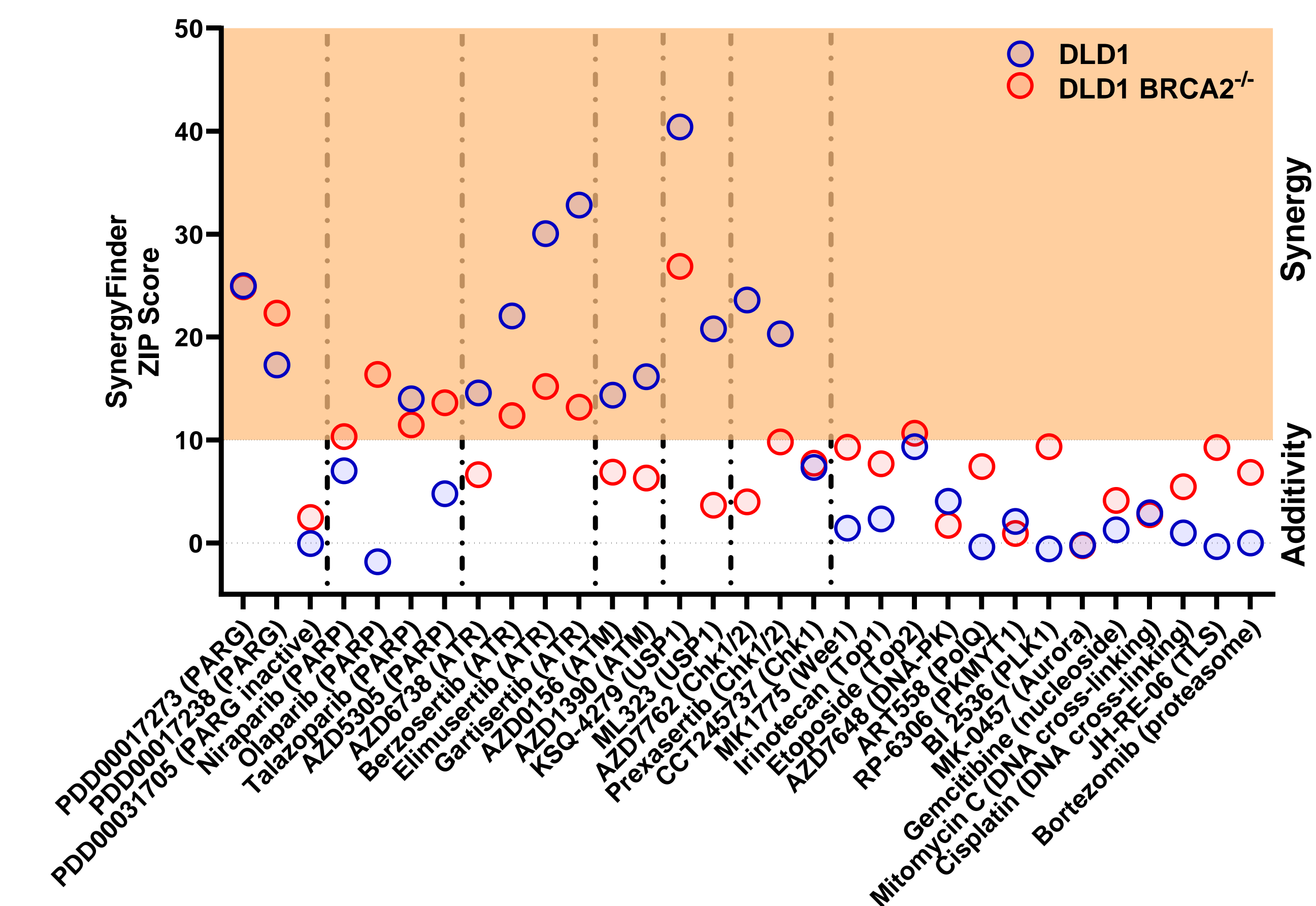
Intracellular staining of chromatin-bound RPA32 in DLD1 and DLD1 BRCA2<sup>-/-</sup> cells treated with 3μM BSM-1516 for 16h. Soluble RPA32 was extracted with detergent prior to fixation and subsequent analysis by flow cytometry.



Western Blot analysis of Ser33-phosphorylated RPA32, an ATR-mediated modification, in DLD1 and DLD1 BRCA2<sup>-/-</sup> cells treated with 3μM BSM-1516 for 24h.

Pharmacological inhibition of FEN1 by BSM-1516 resulted in G2/M cell cycle arrest of BRCA2-deficient DLD1 cells. Analysis of BrdU incorporation and total DNA content was monitored by flow-cytometry.

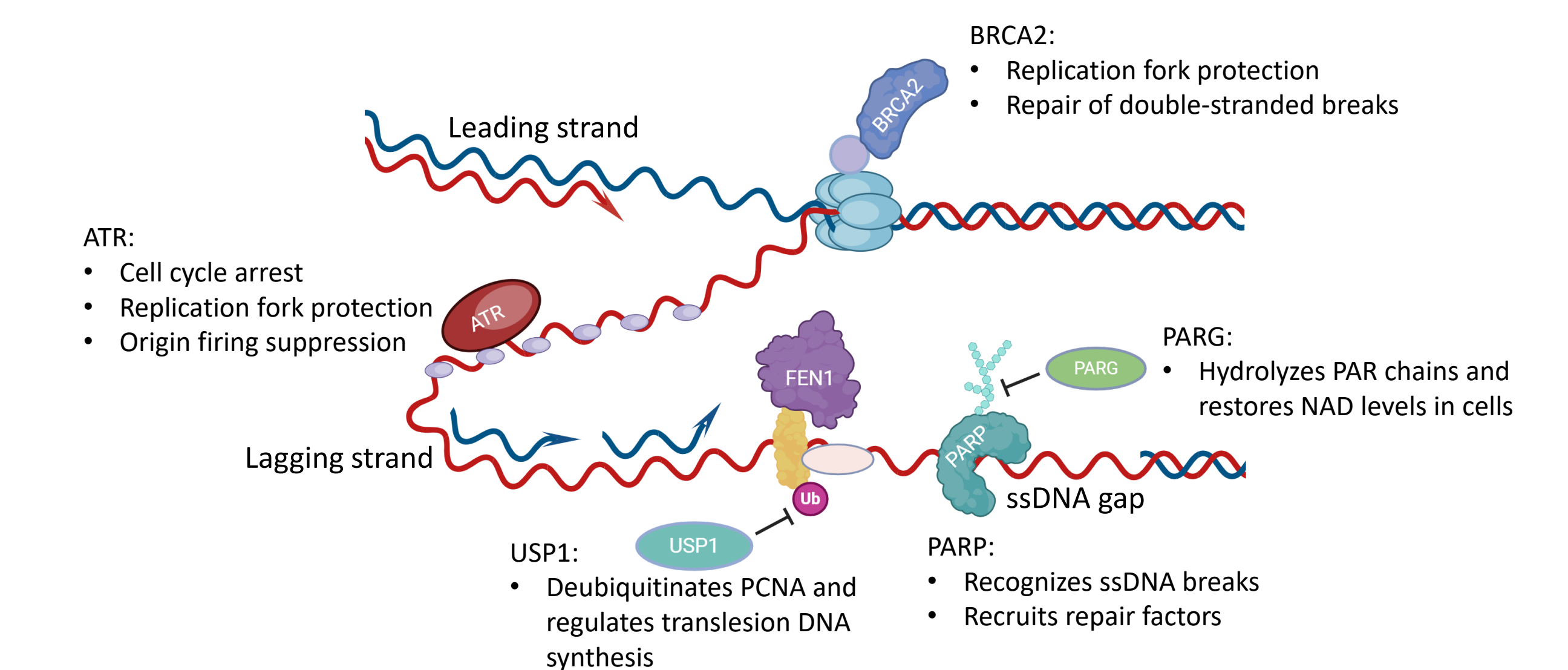
## Drug combination data: BSM-1516 synergizes with inhibitors of USP1, PARG, PARG, ATR and Chk1



To explore BSM-1516 combination potential we performed combination treatment screen with a panel of DNA damage inhibitors and agents. Cell lines were assayed in 6x10 combination half-log dose-response matrix using CellTiter-Glo® 2.0 Cell Viability Assay for 6 days (DLD1) and 9 days (DLD1 BRCA2<sup>-/-</sup>) with media change every 3 days. Synergy scores were calculated using Synergy Finder.

## Summary

- Blacksmith Medicines FBDD approach for metalloenzymes identified a series of cell active FEN1-selective inhibitors that use a novel metal-binding pharmacophore.
- Pharmacologic inhibition of FEN1 is synthetic lethal with defects in HRR and synergizes with inhibitors of USP1, PARG, PARG, ATR and Chk1.
- BSM-1516 exhibits favorable *in vivo* PK properties and its *in vivo* efficacy testing is underway.



## Acknowledgements

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